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drain (Diagram, *c*). The plates are then washed in several changes of water which is drawn from the faucet (Diagram, *s*). Then they can be taken to the light of the room and placed in the fixing bath.

The box as described is a convenient size for use with 4x5 and 5x7 plates. With a 5x7 plate tank it would be well to make the water tank slightly larger. The cost, as estimated by the carpenter who made it, is ten dollars.

Description of Plate I.

Fig. 1. Front view of photographic dark box showing openings for arms with sleeves attached, funnel and water tank at left and door at right.

Fig. 2. End view of the same with door removed to show interior of box. The inlet tube from the water tank and the faucet show above and the drain pan below. The sleeves show at the left.

Fig. 3. Back view of the above showing windows at top and back partly open exposing the red glass and door at the end partly open.

Fig. 4. At work at the photographic dark box.

ELDA R. WALKER.

METHODS OF PREPARING TELEOST EMBRYOS FOR CLASS USE

Some of the most important features of development of the teleost egg, such as discoidal cleavage and the part played by the germ ring in the formation of the embryo, require for their satisfactory demonstration that the entire egg shall be preserved and studied as nearly as possible in its natural form. For this purpose balsam mounts are of little value, for even with the vitelline membrane removed the egg usually collapses in the higher alcohols or in xylol.

For the convenient handling of cleavage stages to be studied as opaque objects I have devised the following method: Pieces of ⅛ inch glass tubing are sealed at one end by holding in a flame; a few eggs fixed in corrosive acetic and preserved in formalin are placed in each tube and the opening plugged with cotton. For these stages it is well to choose an egg having dark yellow yolk; this will aid in the differentiation of the snowy white blastodisc. The eggs may be studied by dropping the tube into a watch glass filled with water and examining them with a lens; or the tube may be held in the hand and examined with the lens. The special advantage of the tube is that of ease of manipulation; one can readily secure any view desired and that without injury to the egg. An additional advantage is that the method allows little trouble from the inevitable mixing of stages that occurs when the material is handled in bulk by students.

For stages with embryonic shield and germ ring the preceding method should be used if the yolk is opaque; but a better plan is to secure the small transparent eggs of the runner *(Ctenolabrus)* and mount them in formalin on a hollow-ground slide, sealing the edges of the cover slip with cement. Staining is not required. In this egg one is able to view either upper or under surface simply by changing the focus.

Michigan State Normal College,
Ypsilanti.
BERTRAM G. SMITH.

PRESERVATION OF BRYOZOA

A method for the preservation of Bryozoa in a well extended condition was worked out at the University of Michigan Biological Station during the summer of 1911.

Care and patience are necessary in the narcotization and killing of Bryozoa.

The following method was used successfully for preserving Cristatella, Plumatella and Fredericella.

Chloretone was used for narcotization, and 3% formalin for killing and subsequent preservation.

The colony was placed in a tube or beaker of convenient size, and covered completely with water. When the lophophores of the individual polypides were well extended, the chloretone solutions were added in the order given below:

1. A few drops of sat. sol. of chloretone in water.
2. 1 part sat. sol. of chloretone to 4 parts of water.
3. 2 parts sat. sol. of chloretone to 3 parts of water.
4. 3 parts sat. sol. of chloretone to 2 parts of water.
5. 4 parts of sat. sol. of chloretone to 1 part of water.
6. Saturated solution of chloretone.